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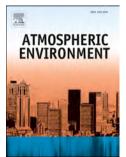
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Air pollutants degrade floral scents and increase insect foraging times

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Abstract

Flowers emit mixtures of scents that mediate plant-insect interactions such as attracting insect pollinators. Because of their volatile nature, however, floral scents readily react with ozone, nitrate radical, and hydroxyl radical. The result of such reactions is the degradation and the chemical modification of scent plumes downwind of floral sources. Large Eddy Simulations (LES) are developed to investigate dispersion and chemical degradation and modification of floral scents due to reactions with ozone, hydroxyl radical, and nitrate radical within the atmospheric surface layer. Impacts on foraging insects are investigated by utilizing a random walk model to simulate insect search behavior. Results indicate that even moderate air pollutant levels (e.g., ozone mixing ratios greater than 60 parts per billion on a per volume basis, ppby) substantially degrade floral volatiles and alter the chemical composition of released floral scents. As a result, insect success rates of locating plumes of floral scents were reduced and foraging times increased in polluted air masses due to considerable degradation and changes in the composition of floral scents. Results also indicate that plant-pollinator interactions could be sensitive to changes in floral scent composition, especially if insects are unable to adapt to the modified *scentscape*. The increase in foraging time could have severe cascading and pernicious impacts on the fitness of foraging insects by reducing the time devoted to other necessary tasks.

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Keywords: Ozone; hydroxyl radical; pollination; pollution; hydrocarbons; fragrances; floral scents

1 1. Introduction

Plants emit a plethora of volatile organic compounds through organs such as roots, leaves, 2 flowers, and fruits (Fuentes et al., 2000). Flowers typically release complex blends of volatile 3 compounds (Knudsen et al., 2006) that give each plant species unique and characteristic 4 fragrances. Although terpenoids, benzenoids, and fatty acid derivatives dominate the floral 5 volatiles, the particular compounds produced by a given species vary among plant groups. 6 For instance, flowers of the mustard plant family (Brassicaceae) typically emit nitrogen-7 containing compounds (Dobson, 2006) while flowers of the dogbane family (Apocynaceae) 8 produce sulfur-containing compounds (Von Helversen et al., 2000). Once biosynthesized, 9 emission rates of floral volatiles depend on biotic and abiotic factors. Biotic influences include 10 flower ontogeny, pollination status, and insect infestation whereas abiotic conditions involve 11 temperature, photosynthetically active radiation, and atmospheric carbon dioxide (CO_2) 12 concentration. Therefore, in downwind areas from sources, ambient concentrations of floral 13 volatiles should exhibit pronounced temporal and spatial variability. Once released into the 14 atmosphere, floral hydrocarbons play key ecological roles such as attraction of pollinators and 15 seed dispersers, defense against herbivores, protection against pathogens, and plant-to-plant 16 signaling (Dötterl and Vereecken, 2010; Wright and Schiestl, 2009). Floral scents provide 17 insect and animal pollinators the necessary cues about location and identity of plant species. 18 Other ecological functions of volatile organic compounds include protection of foliage against 19 abiotic stresses such as high solar radiation loadings and elevated temperature (Sharkey et al.. 20 2001) or oxidative stress (Loreto et al., 2001). 21

One of the least studied aspects of floral volatile composition is the post-emission modifications caused by the reactions with atmospheric oxidants. As floral volatiles are transported by the wind from their sources, they can undergo rapid reactions with air pollutants such as ozone (O_3), hydroxyl radical (OH), and nitrate radical (NO_3). As they traverse landscapes, reactions reduce the strength and the integrity of the chemical cues that floral scents provide

to insects. Their reactions lead to the formation of secondary long-chain hydrocarbon prod-27 ucts, some of which have common chemical characteristics to the parent floral fragrances 28 (McFrederick et al., 2008). The destruction of primary signals and formation of secondary 29 cues are poorly quantified, but are potentially crucial to understanding ecological and evo-30 lutionary conditions for organisms that depend on olfactory cues in polluted environments. 31 Perturbations of scent signals hasten the destruction and also change the identity of scent 32 plumes, both of which can affect the probability of detection by insects. Chemical degrada-33 tion and changes in the composition of floral scents may increase the time that insects need 34 to locate flowers, thereby potentially disrupting processes such as pollination. Flowers could 35 alter the synthesis and the basal emissions of volatiles when exposed to heavy doses of ozone 36 (e.g., > 150 ppbv, Loreto and Schnitzler (2010)) but such influences are not well known. 37

While previous studies examined the influences of habitat fragmentation (Bhattacharya 38 et al., 2003; Montero-Castaño and Vilà, 2012), reductions in native habitat (Goulson et al., 39 2008), pesticide exposure (Whitehorn et al., 2012; Stanley et al., 2015), and diseases (Cameron 40 et al., 2011) on insect communities, only limited attention (Pinto et al., 2007; McFrederick 41 et al., 2008; Blande et al., 2010; Fuentes et al., 2013; Li and Blande, 2015) has been given to 42 investigate the indirect impacts of air pollutants on the plant-insect interactions mediated 43 via floral scents. Changes in composition and reduced amounts of the scent bouquet can lead 44 to insects spending greater amounts of time foraging for food. With the continued changes 45 in OH concentrations (Prinn et al., 2005) and in some cases increases in ground-level O_3 con-46 centrations in the rural atmosphere (Jaffe and Ray, 2007; Cooper et al., 2012; Parrish et al., 47 2013), the modified atmospheric chemistry of floral scents can interfere with their ecological 48 roles. A pressing concern is that the alteration of the natural *scentscape* (McFrederick et al., 49 2009) by air pollutants may be adding to the stresses already contributing to the decline 50 of pollinator populations (Biesmeijer et al., 2006) likely in response to increased pollinator 51 foraging times and decreased pollination efficiency, and exposure to pesticide residues (Stan-52 ley et al., 2015). Despite the potential environmental and economic consequences of the 53 disruption of insect pollination, no information exists to quantitatively evaluate direct and 54

indirect influences of airborne pollutants on the ability of pollinators to sense plant chemical
signals.

Therefore, this study evaluates the hypothesis that *current levels of airborne pollutants* 57 sufficiently alter the composition and reduce the quantity of floral scents in such a manner 58 that they perincipal increase insect foraging times. Several objectives are achieved to 59 evaluate the hypothesis. Large Eddy Simulations (LES) coupled with chemical reactions 60 allow us to determine degradation rates of individual floral volatiles as air parcels carry 61 scents away from sources, exposed to different levels of air pollutants. Also, changes in 62 the composition of floral volatile mixtures are calculated in response to different levels of 63 air pollutants and distances travelled away from scent sources. Finally, foraging times of 64 insects that have different thresholds of floral volatile detection are estimated in response to 65 chemical degradation and modified composition of scent mixtures. A random walk model is 66 developed to estimate insect foraging times as a function of modified scent plumes. 67

68 2. Methods

The understanding of how air pollutants deteriorate interactions between plants and in-69 sects requires knowledge of atmospheric chemistry and animal ecology and physiology. On 70 the atmospheric side, knowledge of air turbulence is required to determine the transport of 71 scents as they traverse the local landscape. Kinetics of floral volatiles released into the at-72 mosphere are necessary to determine how individual compounds are altered by amounts and 73 types of pollutants in the air mass. On the insect side, knowledge is required of the particular 74 floral volatiles or mixtures of compounds that can be distinguished, concentration thresholds 75 required to elicit a response, and the relative importance of scent over other sensory modal-76 ities (e.g., vision) in mitigating the interactions. Currently, no natural system is sufficiently 77 well known to bring together all of the necessary information for reliable predictions in a 78 realistic ecological framework. There is, however, existing theoretical and laboratory work 79 indicating that current air pollutant levels may already be impacting important ecological 80 interactions in natural environments (Fuentes et al., 2013; Farré-Armengol et al., 2016). We, 81

therefore, seek to quantify the impacts of air-pollutant modified scents on plant-insect interactions by overlaying a spatially explicit numerical model of volatile degradation under specified environmental conditions (realistic wind velocities and air pollution regimes) with a foraging model that challenges insects to locate pollutant-modified scent plumes on the landscape.

Large eddy simulations, combined with air chemistry, allow us to determine the turbulent 87 transport and chemical reactions of floral volatiles. The model predicts the time evolution 88 of scent plumes under the influences of varying levels of turbulence mixing and chemical 89 reactions at small spatial scales, thereby providing detailed information of floral scent plume 90 composition with high temporal and spatial resolution (Figure 1). Therefore, in the absence 91 of detailed information from field observations, LES outputs are the most reliable source of 92 information for studying transport and chemical degradation of scent plumes. A random 93 walk model is developed and applied to compute foraging times based on insect detection 94 threshold of floral scents and modified *scentscape*. 95

96 2.1. Description of the LES

The LES model used in this study to determine dispersion of floral volatiles by the wind is described in detail elsewhere (Chamecki et al., 2009) and only a brief description is included here. In the LES, the filtered three-dimensional wind field ($\widetilde{\mathbf{u}}$) is determined from the solution of the filtered momentum and mass conservation equations which in vectorial form are expressed as (∇ is the vector differential operator)

$$\nabla \cdot \widetilde{\mathbf{u}} = 0 \tag{1}$$

$$\frac{\partial \widetilde{\mathbf{u}}}{\partial t} + (\widetilde{\mathbf{u}} \cdot \nabla) \widetilde{\mathbf{u}} = -\frac{1}{\rho} \nabla \widetilde{p} - \nabla \cdot \boldsymbol{\tau}_{\text{sgs}}.$$
(2)

In equation (2), the term $-(1/\rho)\nabla \tilde{p}$ is the filtered pressure gradient force and τ_{sgs} is the subgrid-scale (SGS) stress tensor representing the effect of the scales smaller than the LES grid size on the resolved wind field. Viscous effects are neglected on the basis of the large Reynolds number of the flows considered in this study. Additional filtered advection-

diffusion-reaction equations were included in the LES model to represent the temporal variations of floral volatile concentrations $(\partial [\widetilde{\chi_i}]/\partial t)$

$$\frac{\partial[\widetilde{\chi_i}]}{\partial t} + \nabla \cdot \left(\widetilde{\mathbf{u}}[\widetilde{\chi_i}]\right) = -\nabla \cdot \boldsymbol{\pi}_{\chi_i} + Q_{\mathrm{src},i} - k_{\mathrm{O}_3}[\widetilde{\chi_i}][\mathrm{O}_3] - k_{\mathrm{HO}}[\widetilde{\chi_i}][\mathrm{HO}] - k_{\mathrm{NO}_3}[\widetilde{\chi_i}][\mathrm{NO}_3].$$
(3)

In equation (3), $[\chi_i]$ is the concentration of the *i*th floral volatile, π_{χ_i} is the SGS floral 108 volatile flux representing the mixing caused by turbulence scales not explicitly included in 109 the simulation, $Q_{\text{src},i}$ represents the emission of scent from flower patches, and the three last 110 terms on the right-hand side of equation (3) represent the loss of $[\widetilde{\chi_i}]$ due to reaction with 111 O_3 , OH, and NO₃ with reaction rate coefficients k_{O_3} , k_{HO} , and k_{NO_3} . To close the set of 112 equations, the SGS momentum flux is parameterized using the Lagrangian scale-dependent 113 dynamic Smagorinsky model (Bou-Zeid et al., 2005). The SGS floral volatile fluxes are then 114 parameterized using the resulting SGS eddy viscosity and the SGS Schmidt number (Sc_{τ}) 115 which in this study is assumed to be 0.8. 116

The momentum equations are discretized using a fully de-aliased, pseudo-spectral ap-117 proach in the horizontal directions and a second-order centered finite-difference scheme in 118 the vertical direction. Model lateral boundary conditions are periodic. A no-stress boundary 119 condition is imposed at the top boundary and an equilibrium wall-model (Bou-Zeid et al., 120 2005) is used to parameterize the bottom boundary conditions. The advection-diffusion-121 reaction equation (3) is discretized following a finite-volume method, with the bounded 122 third-order upwind advection scheme SMART (Gaskell and Lau, 1988). The coupling be-123 tween the pseudo-spectral discretization for momentum and the finite volume discretization 124 for reactive scalars uses a conservative interpolation scheme (Chamecki et al., 2008). The 125 lateral boundary conditions for the floral volatiles are specified as zero values for inflow and 126 zero gradients for outflow boundary conditions and zero-flux conditions are specified at the 127 top and bottom boundaries. The time integrations of wind velocity and floral volatile con-128 centrations are carried out using the second-order Adams–Bashforth scheme (Peyret and 129 Taylor, 1983). The flow is driven by an imposed mean pressure gradient, which is adjusted 130 to generate different wind conditions (corresponding to friction velocity $u_* = 0.1 \text{ m } s^{-1}$ and 131

 $u_* = 0.2 \text{ m } s^{-1}$). Neutral thermal stratification is assumed in all simulations. The resulting vertical variations of averaged zonal wind speed (\overline{u}) and standard deviation of the vertical velocity (σ_w) are included in Figure 2 for the assumed u_* values.

The simulation domain consists of a volume of $1000 \,\mathrm{m} \times 350 \,\mathrm{m} \times 50 \,\mathrm{m} \,(L_x \times L_y \times L_z)$, 135 covered by a sparse vegetation of height h = 1 m (represented by a constant surface roughness 136 $z_0 = 0.05 \,\mathrm{m}$). This domain is discretized using $400 \times 140 \times 100$ grid points, resulting in a 137 grid resolution of $2.5 \,\mathrm{m} \times 2.5 \,\mathrm{m} \times 0.5 \,\mathrm{m}$. Flower patches are randomly distributed within an 138 area of $180 \,\mathrm{m} \times 250 \,\mathrm{m}$ near the upwind edge of the domain (Figure 1). Five hundred flower 139 patches (with $2.5 \text{ m} \times 2.5 \text{ m}$ each) are included in the simulations. Flowers are asummed to 140 emit blends of volatiles comprised of β -caryophyllene, β -ocimene, β -myrcene, linalool, and 141 α -pinene (i.e., i = 1, ..., 5 in Equation 3 and Table 1). The chosen volatiles (Table 1) are 142 common components of floral fragrances and play numerous ecological functions, including 143 attracting pollinators (Byers et al., 2014), herbivores (Mayer et al., 2008), and parasitoids 144 of herbivores (Ozawa et al., 2008). Herbivore feeding on plants enhances production of β -145 myrcene as a way of attracting parasitoids to control the herbivores (Van Poecke et al., 146 2001). Basal emission rates $(E_{s,i})$ for a nominal temperature $T_s = 303$ K (Table 1) for each 147 floral volatile are determined from experimental data (Wright et al., 2005), assuming a leaf 148 area index equal to 2. Effective emission rates (E_i) corresponding to a flower temperature 149 of $T = 298 \,\mathrm{K}$ are used in the simulation, and are determined employing equation (4) 150

$$E_i(T) = E_{s,i} \exp\left[\beta(T - T_s)\right],\tag{4}$$

where β is a constant taken as 0.06 K^{-1} . The floral volatile emission per unit volume required in equation (3) is then calculated as $Q_{\text{src},i} = E_i(T)/dz$, where dz is the vertical grid spacing used in the simulation. Each simulation is executed for 2 hours with a time step dt = 0.1 s. Data analyses are performed for the final hour of the simulations, when gas concentration fields are statistically stationary.

156 2.2. Model of insect foraging flight

Random walk models are frequently used to represent the foraging behavior of animals
 (Codling et al., 2008). Typically, resources are represented as points distributed throughout

| Index | Species | Emission rate (E_s) | K_{O_3} | K _{OH} | $K_{\rm NO_3}$ |
|-------|------------------------|-----------------------------|---------------------------------------|---|---|
| | | (nmoles $m^{-2} min^{-1}$) | $(cm^3 molec^{-1} s^{-1})^{\ddagger}$ | $(\rm cm^3 \ molec^{-1} \ s^{-1})^{\ddagger}$ | $(\rm cm^3 \ molec^{-1} \ s^{-1})^{\ddagger}$ |
| 1 | β -Caryophyllene | 15 | 1.10×10^{-14} | 2.00×10^{-10} | 1.90×10^{-11} |
| 2 | β -Ocimene | 82 | 5.40×10^{-16} | 2.52×10^{-10} | 2.20×10^{-11} |
| 3 | β -Myrcene | 25 | 4.70×10^{-16} | 2.13×10^{-10} | 1.27×10^{-11} |
| 4 | Linalool | 6.5 | 4.30×10^{-16} | 1.59×10^{-10} | 1.12×10^{-11} |
| 5 | α -Pinene | 10 | 8.09×10^{-17} | 5.33×10^{-11} | 6.16×10^{-12} |

Table 1: Floral volatiles, emission rates, and reaction rate coefficients used in the model simulations. ‡: Values of reaction rate coefficients were obtained from Atkinson et al. (1999).

the landscape and random walks are then used to represent the movement of animals as they 159 survey the surroundings in search of resources. A radius of detection is typically assigned to 160 the forager and once a resource is within this radius then the search is considered success-161 ful. In this typical approach, the existence of chemical cues such as scent plumes is either 162 neglected or is included in the detection radius of the insect. Therefore, external factors 163 (such as turbulence and air pollutant levels) that influence composition and spatial extent 164 of the floral scent plumes cannot be studied. In the present work, we address this issue by 165 explicitly representing the spatial extent of floral scent plumes that are generated from the 166 LES model outputs. 167

From all the possible random walk models, Lévy walks have received significant attention 168 (Viswanathan et al., 1999; Reynolds and Rhodes, 2009; Reynolds, 2010). Under some specific 169 circumstances, Lévy walks represent optimal search strategies (Viswanathan et al., 1999; 170 Raposo et al., 2009). More recently, the idea of composite searches (Plank and James, 2008) 171 in which insects invest foraging times in resource rich areas has gained attention. Basically, 172 insects move back and forth between two strategies: an intensive search is adopted in regions 173 rich in resources and an extensive foraging strategy is adopted elsewhere. Thus, insects 174 first use extensive searches to locate resources, and then they alternate to intensive searches 175 within areas with resources. If searches are not successful then insects move back to extensive 176

foraging to cover larger areas in search of other resource rich areas. Even in composite search
strategies, Lévy walks seem to be the most profitable approach (Reynolds and Bartumeus,
2009). Despite recent criticism regarding the optimality of Lévy walks as a foraging strategy
(Pyke, 2015), the approach is still useful as an idealized model to investigate the effects of
environmental characteristics on insect foraging.

¹⁸² We adopt the Lévy walk to represent insects searching for floral scent plumes. In the ¹⁸³ context of composite searches, this can be viewed as the extensive strategy of locating the ¹⁸⁴ first resource location within an unknown landscape (Reynolds and Bartumeus, 2009; Nolting ¹⁸⁵ et al., 2015). In the model, insect flights are determined by a step length (l) and a horizontal ¹⁸⁶ angle (θ). Step lengths are drawn from a power-law distribution given by

$$P(l) = \left(\frac{l}{l_0}\right)^{-\mu}.$$
(5)

In equation (5), l_0 is the minimum step length and μ is a parameter whose value ranges from 187 1 to 3. Note that $\mu = 3$ corresponds to Brownian motion and $\mu = 2$ is a super-diffusive Lévy 188 walk. The model with $\mu = 2$ represents optimal search strategies (Viswanathan et al., 1999) 189 and it is used in the current study. Horizontal angles are drawn from a uniform distribution. 190 Two source distributions representing landscapes with different amounts of resources are 191 investigated. In the first case, only one simulated plume is included, representing a landscape 192 with scarce resources. Insects are released from the downwind corner of the simulation 193 domain ($x = 1000 \,\mathrm{m}$ and $y = -175 \,\mathrm{m}$), corresponding to a distance of nearly 1 km from 194 the center of the flower patch (Figure 1, top panel). A landscape rich in resources is also 195 considered. For that purpose, eight copies of the scent plumes produced by the LES are 196 placed in a circular pattern around the insect release location (Figure 1, bottom panel). In 197 both cases, the initial height (z) of the insects is set to 1 m, and this value is kept constant 198 during the insect flights. Each insect flight step is broken into substeps of size l_0 ($l_0 = 1 \text{ m}$ 199 is used here) and, after each substep, the local floral scent concentration is compared to the 200 insect detection threshold for a given compound to decide whether the insect is capable of 201 recognizing the scent plume at that location. If the gas concentration is greater than the 202 detection threshold, the flight ends and the insect is considered successful in locating the 203

scent plume. In the model, it is assumed that insects fly at a constant speed of 2 m s^{-1} and terminate their flights if the scent plume is not found within 3 hours. For each scenario (see description in Section 2.3), 18 instantaneous LES-generated scent plumes are considered. For each plume, a total of 5,000 insect flights are considered, and statistics are computed over the accumulated 90,000 flights. Simulations are performed for detection thresholds varying from 0.1 pptv to 25 pptv. These detection thresholds are within the lowest-observed responses of insects (Dusenbery, 1992).

211 2.3. Pollution scenarios

Six scenarios (Table 2) are considered to determine changes in scents in response to 212 varying levels of air pollutants as air parcels travel away from flowers. The first scenario 213 (Sc0) considers no air pollutants to determine the spatial variation of floral volatiles away 214 from sources in response to turbulent transport and to serve as the basis for comparison 215 with the rest of scenarios. The second scenario (Sc1) includes 20 ppbv of O_3 , 0.02 part per 216 trillion on a volume basis (pptv) $(5.0 \times 10^5 \text{ radicals cm}^{-3})$ of OH, and 0 pptv of NO₃. This 217 scenario may be viewed as the prevailing O_3 levels during pre-industrial times (Marenco 218 et al., 1994), with limited photochemical activity (and hence low OH levels) and no nitrogen 219 oxides to generate NO₃. The third scenario (Sc2) involves the levels of 0.2 pptv (5.0×10^6 220 radicals cm^{-3}), 40 ppbv, and 0 pptv of OH, O₃, and NO₃, respectively. The fourth scenario 221 (Sc3) uses the levels of 0.3 pptv $(1.0 \times 10^7 \text{ radicals cm}^{-3})$, 60 ppbv, and 1 pptv for HO, O₃, 222 and NO₃, respectively. The fifth scenario (Sc4) involves the levels of 0.40 pptv (2.0×10^7) 223 radicals cm^{-3}), 80 ppbv, and 2 pptv for OH, O₃, and NO₃, respectively. The last scenario 224 (Sc5) involes the levels of 0.80 pptv $(2.0 \times 10^7 \text{ radicals cm}^{-3})$, 120 ppbv, and 5 pptv for 225 OH, O_3 , and NO_3 , respectively. We choose these last two scenarios to reflect the broad 226 ranges in air pollutant deviations from pre-industrial times (Marenco et al., 1994; Prinn 227 et al., 2005) to current summertime conditions in regions such as the eastern United States 228 where maximum O_3 levels can sometimes exceed 120 ppbv (Fiore et al., 2002). Pollutants 229 are assumed to logarithmically vary with altitude, and the values listed here are specified 230 at a height $z = 1.5 \,\mathrm{m}$ (the value at the top of the domain is twice the value at 1.5 m). For 231

- 232 simplicity and convenience, for a given scenario, the ambient mixing ratios of OH, O₃, and
- $_{233}$ NO₃ remain constant in time and space throughout the course of model simulations.

 Table 2: Ambient mixing ratios of the ozone, hydroxyl radical, and nitrate radical included in the scenarios of the numerical simulations.

| Scenario | O_3 | НО | NO ₃ |
|----------|--------|--------|-----------------|
| | (ppbv) | (pptv) | (pptv) |
| Sc0 | 0 | 0.00 | 0 |
| Sc1 | 20 | 0.02 | 0 |
| Sc2 | 40 | 0.20 | 0 |
| Sc3 | 60 | 0.30 | 1 |
| Sc4 | 80 | 0.40 | 2 |
| Sc5 | 120 | 0.80 | 5 |

234 2.4. Data analysis techniques

In total, twelve LES runs are done to include the six air pollutant scenarios under the influences of two turbulence levels. Average floral volatile concentrations are calculated to determine the chemical degradation of scent plumes in response to varying levels of air pollutants. For each floral scent (i = 1, ..., 5, Table 1) and scenario ($Sc_j, j = 0, ..., 5$), temporal averages are performed (for periods defined from initial time (T_0) to some later time (T_f)) and crosswind direction to yield mean gas concentration as a function of x and z

$$\overline{[\chi_i]}_{Sc_j}(x,z) = \frac{1}{L_y(T_f - T_0)} \int_{T_0}^{T_f} \int_{-L_y/2}^{L_y/2} [\widetilde{\chi_i}]_{Sc_j}(x,y,z,t) dy dt.$$
(6)

Resulting average scent concentrations are reported in two ways. First, to illustrate the effects of air pollutants on scent concentrations downwind from sources, gas concentrations are normalized to concentrations for the scenario without air pollutants ($\overline{[\chi_i]}_{N_j}(x,z)$) and are determined using Equation (7) for scent species i = 1, ..., 5.

$$\overline{\left[\chi_{i}\right]}_{N_{j}}\left(x,z\right) = \frac{\overline{\left[\chi_{i}\right]}_{\mathrm{Sc}_{j}}\left(x,z\right)}{\overline{\left[\chi_{i}\right]}_{\mathrm{Sc}_{0}}\left(x,z\right)}$$
(7)

Second, to emphasize the effects of air pollutants on the changes in the composition of scent plumes, fractional composition $(\overline{[\chi_i]}_{F_j}(x,z))$ of scent plumes are computed using Equation (8) for each scenario.

$$\overline{[\chi_i]}_{F_j}(x,z) = \frac{\overline{[\chi_i]}_{\mathrm{Sc}_j}(x,z)}{\sum_{k=1}^5 \overline{[\chi_k]}_{\mathrm{Sc}_j}(x,z)}$$
(8)

Only values for the height of insect flight (z = 1.0 m) are reported below (i.e., $\overline{[\chi_i]}_{N_j}(x, 1.0 \text{ m})$, $\overline{[\chi_i]}_{F_j}(x, 1.0 \text{ m})$). For the fractional composition, values are reported at the discrete downwind distances (x) of 200, 300, 400, and 800 m.

For insect foraging times, statistics are calculated for each flower volatile and for each air pollution scenario individually. Calculations are done separately for landscapes scarce and rich in scent resources. Summary statistics are presented for the discovery of floral plumes based on three detection thresholds. The cumulative distributions of insect discovery times of floral plumes and the proportion of insects discovering floral plumes within 3 hours are presented.

257 3. Results

258 3.1. Chemical degradation of floral volatiles

The chemical degradation of floral scents depends on the reactivity of volatiles and am-259 bient levels of air pollutants. For the chosen air pollutant levels, floral chemical species 260 exhibit a broad range of lifetimes (for the reaction with O_3 , the individual floral scent (χ_i) 261 lifetime is defined as $\tau_{\chi_i,O_3} = (K_{\chi_i,O_3}[O_3])^{-1}$; similar expressions can be defined for $\tau_{\chi_i,OH}$ 262 and τ_{χ_i, NO_3}). Concerning the O₃ reactivity, β -caryophyllene is the most reactive scent and 263 its lifetime decreases from 12 to 2 minutes for O_3 levels between 5 and 120 ppbv (Figure 264 3). Linalool, β -ocimene, and β -myrcene have an intermediate reactivity with respect to O₃ 265 and their lifetimes vary from 200 to 10 minutes when O_3 changes from 5 to 120 ppbv. In 266 these simulations, α -pinene is the least reactive with O₃ and its lifetime changes from 30 to 1 267 hours for O_3 levels between 5 and 120 ppbv (Figure 3). With regard to the OH reactivity, β -268 ocimene, β -myrcene, and β -caryophyllene are the most short lived scents and their lifetimes 269 range from 50 hours to 5 minutes when OH changes from 0.001 to 0.8 pptv. Concerning 270

the NO₃ reactivity, β -ocimene and β -caryophyllene are the most reactive species and their lifetimes range from 18 hours to 5 minutes as NO₃ varies from 0.01 to 5 pptv. In polluted air masses (i.e., [OH] > 0.3 pptv), all floral scents have lifetimes of few (< 30) minutes (Figure 3).

Source strength of floral volatiles, levels of air turbulence, and rates of chemical reac-275 tions dictate the three-dimensional characteristics of the scents downwind from sources. For 276 example, Figure 4 displays iso-surfaces of time averaged scent mixing ratios in the three-277 dimensional space (i.e., a surface formed by all the points with a chosen constant gas mixing 278 ratio value) downwind from the chosen floral patches. Blue iso-surfaces represent mixing 279 ratios of 0.6 pptv for myrcene and β -caryophyllene (under the influences of scenarios Sc0, 280 Sc3, and Sc5), indicating that all the points inside the blue surface have scent levels above 281 0.6 pptv and all the points outside the surface have scent mixing ratios below 0.6 pptv. 282 The volume inside the blue iso-surface represents the portion of the plume that an insect 283 with a threshold detection of 0.6 pptv can detect. Similarly, the volume inside the purple 284 iso-surfaces represents the portion of the scent plume that insects with a detection threshold 285 of 4.8 pptv can detect. 286

Ambient scent distribution downwind from sources is inversely related to turbulence 287 levels (Figure 4). For the same floral volatile emission rates, scent levels are higher under 288 the influence of low air turbulence (this feature is clearly noticeable on the purple plumes). 289 This effect is due to the reduced dilution rates of scents caused by lower mean wind speeds 290 associated with the low turbulence (Figure 2). Also, under the influences of the two adopted 291 levels of turbulence and no chemistry, relatively elevated gas mixing ratios prevailed across 292 the landscape, with scent levels above 3.2 pptv extending as far as 300 m away from flowers 293 (note that the source ends at $x = 200 \,\mathrm{m}$, so the location at $x = 500 \,\mathrm{m}$ is 300 m downwind 294 from the flower patch). Sufficient levels (>0.4 pptv) of β -myrcene (Figure 4A) and β -295 caryophyllene (Figure 4B) reached about 800 m downwind from floral volatile sources. Such 296 scent spatial distribution patterns and abundances were deemed adequate for most insects to 297 locate the floral scent plumes (Dusenbery, 1992). In effect, the resulting spatial distribution 298

of floral scents provided information on the effective plume "footprint" of a particular floral patch. For polluted conditions (scenarios Sc3 and Sc5), the plumes of floral volatiles such as β -myrcene, with medium reactivity, could only reach 400 m downwind from sources (Figure 4). For the highly reactive compounds such as β -caryophyllene, sufficient levels were only found near flowers. Therefore, in polluted air masses, the "volume" of scent plumes is drastically reduced due to the chemical reactions, thereby making it harder for insects to locate food.

Air pollutant levels and reactivity of floral scents exert control on the spatial extent of 306 for floral volatile transported away from sources. Horizontal cross sections (Figure 5) provide the 307 spatial distribution of floral scents downwind from sources under the influences of different 308 levels of air pollutants. To some extent, they represent the *scentscape* that insects can 309 detect while foraging for food resources. The unpolluted case (Sc0) serves as a basis for 310 comparison and shows similar spatial distributions of β -myrcene and β -caryophyllene (see 311 color scales, Figure 5), confirming that the difference in source strength between the two 312 compounds (about 60 %, see Table 1) is not the dominant factor in determining their spatial 313 distributions. Under moderate levels of air pollutant (Sc3), significant levels of β -myrcene 314 $(\sim 1 \text{ pptv})$ can reach about 800 m downwind from floral sources whereas similar levels of 315 β -caryophyllene reach at most 100 m downwind from sources. The most dramatic effect 316 occurs in polluted air masses (Sc6) when plumes of β -myrcene remain within 400 m of scent 317 sources; in contrast, plumes of β -caryophyllene can only be found directly above the floral 318 patches. 319

While foraging for resources most insects use non-directional sensory cues and instantaneous plumes of scents to identify areas that warrant searches (Nolting et al., 2015; Bell, 1990). Instantaneous scent plumes provide information to allow foraging insects to directly orient themselves to habitat resources and determine the appropriate search strategy (i.e., intensive or extensive search modes). Time-averaged plumes (Figure 5) differ substantially compared to instantaneous ones (Figure 6). Whereas time-averaged scent plumes provide relatively smooth scent concentration gradients along horizontal transects, instantaneous flo-

ral fragrance plumes reveal greater heterogeneity in the spatial distribution of gases. Some 327 areas can be almost devoid of floral scents while the contiguous surroundings exhibit high 328 loading of scents (e.g., scenarios Sc3 and Sc5 for β -caryophyllene, Figure 6). The character-329 istic features in the spatially heterogeneous gas distribution in instantaneous plumes result 330 in response to erratic turbulent fluctuations that allow for the rapid transport of gases. In 331 the instantaneous plumes, the intensity of the spatiotemporal scent fluctuations increases 332 with air turbulence (data not shown). Consequently, at any given location and time, scent 333 concentrations within plumes can erratically and substantially vary compared to the time-334 averaged plumes (compare Figure 5 with Figure 6). These results (Figure 6) suggest that 335 foraging insects likely use olfactory information provided by instantaneous plumes to influ-336 ence search behavior at the habitat level that can impact search strategies at the patch, and 337 search at the patch can affect foraging for individual resources (Bell, 1990). 338

Ambient scent levels downwind from sources depend on rates of chemical destruction 339 and turbulence intensity. Under the influences of the assumed air turbulence conditions (u_*) 340 = 0.1 m s⁻¹ and $u_* = 0.2$ m s⁻¹), the normalized time-averaged concentrations of floral 341 volatiles (i.e., $\overline{[\chi_i]}_{N_j}$ for floral scents i = 1, ..., 5 and air pollution scenarios j = 1, ..., 5, see 342 Methods Section) provide a qualitative measure of the influences of chemical reactions on 343 the distances traveled by scent plumes away from sources. Compounds such as β -myrcene 344 and β -caryophyllene can exhibit different abundances along transects from downwind of 345 sources in response to the reactivities of floral volatiles (Figure 7). For example, under 346 the influences of low air pollutant concentrations (Sc1), only 75 to 80% of the emitted β -347 myrcene reaches 800 m away from floral sources, when $u_* = 0.1 \text{ m s}^{-1}$ and $u_* = 0.2 \text{ m s}^{-1}$, 348 respectively. In contrast, only 5% of the emitted β -caryophyllene reaches 800 m when $u_* =$ 349 0.1 m s^{-1} and 400 m away from sources when $u_* = 0.2 \text{ m s}^{-1}$. For highly polluted conditions 350 (Sc6), only approximately 25% of the original β -myrcene remains within distances of 100 m 351 downwind from floral sources (Figure 7). As air pollutant concentrations increase, amounts 352 of scents exponentially decrease with distance away from floral sources. Based on these 353 results (Figure 7), it is concluded that reductions in the amounts of floral scents transported 354

away from sources can impair the ability of insects locating floral scents due to the reductions
of olfactory cues below the insect detection thresholds.

357 3.2. Modification of floral volatile mixtures

Generalist insects such as honeybees respond to floral bouquets instead of a specific scent 358 (Wright et al., 2005; Raguso, 2008). Also, generalist insects respond to compound blends but 359 detect ratios as indicators to judge potential success of locating resources (Raguso, 2008). 360 Because of the different reaction rates for each floral volatiles, however, the composition of 361 the floral scents can change as they react with air pollutants, thereby altering the floral bou-362 quet. To show the potential changes in the ratios of these components, LES results were used 363 to track the composition of scent plumes as they traveled away from from sources. Figure 364 8 depicts the differences between the floral composition at the source and the composition 365 of scent plumes at several downwind distances from sources. While the initial composition 366 of the scent was comprised of 60 % β -ocimene, 20 % β -myrcene, 8 % β -caryophyllene, 7 % 367 β -pinene, and 5 % linalool at the source, it is evident (Figure 8) the fragrance composition 368 changed substantially as air parcels traveled downwind in polluted conditions. For instance, 369 at 800 m away from sources, β -pinene, rather than β -ocimene, became the predominant 370 compound in the floral mixture, due to the slower reaction rates. Even by the short traveled 371 distance of 250 m, the floral bouquet did not include β -caryophyllene due to its destruction 372 via chemical reactions. Therefore, the results indicate that the insects associated with origi-373 nal floral volatile blends would become increasingly 'confused' and unable to distinguish the 374 appropriate composition of the volatile mixture away from the source under polluted con-375 ditions. The addition of pollutants, however, makes this task of identifying floral mixtures 376 more laborious as the scent composition changes much more rapidly with increasing levels 377 of oxidants. 378

For those insects that have evolved to rely upon the unique signature of a specific flower, a change in the composition of these scents could be troubling, depending upon how sensitive their antennae are to detecting particular compounds Skiri et al. (2005) and their tolerance for shifts in relative concentration among plume components. Many insect species hone

in on specific scents because it increases their likelihood of being consistently rewarded. 383 If particular flowers are proven to be reliable sources of nectar for foraging insects then 384 they can train themselves to that specific scent, what is known as flower constancy (Gegear 385 and Laverty, 2005). In polluted conditions, however, the plume of floral volatiles may no 386 longer be recognized as such because its composition no longer corresponds to the scent 387 that insects associate with a particular flower. Moreover, because plant odors mediate both 388 mutualistic and antagonistic relationships between plants and insects, a disruption in the 389 communication system could either help or hurt plants depending on which insects are most 390 affected. In cases where antagonists and mutualists respond to different volatiles from the 391 same plant (e.g., Andrews et al. (2007)), the outcome will depend partly on the reactivity 392 of the different compounds. More generally, however, mutualistic relationships may be more 393 sensitive to disruption because they typically rely on repeated tracking of scent to source 394 (e.g., pollinators and natural enemies searching for new resources) whereas antagonists such 395 as herbivores may find sufficient resources at a single resource once discovered and rely less 396 on long-distance plant signals from that point forward. 397

398 3.3. Potential impact on insect success and foraging times

Air pollutants, type of hydrocarbons, detection thresholds of insects, and number of floral 399 patches on the landscape impact insect discovery times of floral sources. These effects are 400 illustrated in Figure 9, where the cumulative number of insects that locate the scent plume is 401 displayed as a function of foraging time (up to the limit of 3 hours). Each panel corresponds 402 to a combination of hydrocarbon used by insects and corresponding insect detection threshold 403 limit, and results under different pollution scenarios are compared for landscapes scarce and 404 rich in resources (solid and dashed lines, respectively). By itself, detection threshold sets 405 severe constraints on how far away scent plumes can be detected when transported downwind 406 from sources. When there is only a single floral patch on the landscape (Figure 9 – solid 407 lines), the proportion of foragers that locate the scent within 180 minutes in the unpolluted 408 environment decreases from about 80 % with a detection threshold of 0.1 pptv (Figure 9A-C) 409 to approximately 20 % in an unpolluted environment with a detection threshold of 3.9 pptv 410

(Figures 9G-I). These results are due to the fact that insects with low olfactory detection
threshold can detect greater foraging areas in response to greater scent loadings (Figure 6).
Therefore, olfactory detection threshold values explain the broad computed foraging times
for the investigated scents (Figure 9).

The reactivity of the hydrocarbon being modeled strongly influences the effect of air 415 pollutants on scent plume detectability. For the most reactive floral volatile, β -caryophyllene, 416 even modest levels of air pollutants (e.g., $[O_3] = 20$ ppbv, [OH] = 0.02 pptv) can have 417 dramatic effects on both discovery times and probability of discovery within 180 minutes. 418 thereby reducing the likelihood of discovery by 65 % and increasing the discovery time from 419 about 10 minutes to approximately 180 minutes for the first 20 % of foragers to locate 420 the scent plume (Figures 9C and F). As air pollutant levels increase, the size of the scent 421 plume diminishes (Figure 6) which increases the insect foraging times. For a hydrocarbon 422 of intermediate reactivity such as β -myrcene, the plume discovery can broadly differ across 423 all pollutant levels (Figures 9B and E), while for α -pinene, the least reactive hydrocarbon 424 included in the foraging model, a notable effect of air pollutants is only observed at an 425 intermediate level of detectability (i.e., 0.6 pptv, Figure 9D). Again, size of scent plumes and 426 characteristic features of spatial gas distribution (Figure 6) impact the insect discovery times 427 of flower patches. Number of floral patches present throughout the landscape also influences 428 the outcome of foraging trials. As floral patches become prevailingly common (Figure 9 – 429 dashed lines), most foragers encounter scent plumes within 180 minutes. At the extreme, 430 over 50 % of foragers encounter scent plumes within 15 minutes while searching for the 431 most reactive compound in unpolluted environments, but similar numbers of foragers take 432 about 60 minutes for discovery of scent plumes under modest pollution levels (Figure 9F). 433 Therefore, these results indicate that floral scent reactivity, pollutant levels, and detection 434 thresholds all strongly mediate the ability of insects to locate scent plumes. 435

In our foraging model, insects searched for a particular scent in the landscape, following it when it was encountered above a threshold concentration. Because insects are capable of associative learning (Hollis and Guillette, 2015), they could potentially learn to follow

the altered plume to the original plume and reach the reward. Second order conditioning 439 (associating one stimulus with another stimulus) has been demonstrated in the laboratory 440 with honeybees exposed consecutively to two distinct odors (Hussaini et al., 2007). It would 441 be challenging, however, for an insect to learn distinct stimuli under polluted conditions as 442 the odor would be continuously changing with distance and with diurnal patterns of pollutant 443 concentrations. While we cannot currently evaluate the likelihood of an insect learning to 444 track the altered plume, we recognize this effect is an interesting study area with broad 445 implications for insect response to environmental change. 446

447 4. Summary and Conclusions

This study concludes that even moderate air pollutant levels (e.g., $[O_3] > 60$ ppbv) sub-448 stantially degrade the chemical constituents of released floral scents. The chemical degrada-449 tion of floral volatiles contributes to substantial reductions in the concentrations of scents as 450 they are transported away from sources. Decreases in scent abundances away from sources 451 show steeper decreases with increasing levels of air pollutants. In the most polluted air 452 masses ([O₃] = 120 ppbv), compounds such as β -ocimene decrease to 25 % of the original 453 abundance within a downwind distance of 100 m from sources whereas β -caryophyllene re-454 tains only 10 % of the original amount within 50 m of sources. In all scenarios examined, 455 concentrations of the studied floral volatiles substantially decrease in response to increasing 456 levels of air pollutants and rates of scent destruction depend on gas reactivity with respect 457 to O_3 , OH, and NO_3 . Because individual components of floral scents degrade at different 458 rates, the scent itself changes as a function of reactant concentrations and time evolution 459 of the scent plumes. At downwind distances of 800 m, the least reactive compounds (e.g., 460 α -pinene) dominate the composition of scents whereas the more reactive compounds (e.g., 461 β -caryophyllene) are chemically destroyed. 462

The likelihood of foraging insects discovering a particular flower patch depends on air pollutant levels, antennal sensitivity, and the component(s) of the floral scent that the insects recognize and follow. Taken together, these variables determine the perceptual footprint of scent plumes, and the size of that footprint determines both the speed and the likelihood of

patch discovery. Our simulations indicate that air pollution will have the greatest impact
on patch discovery time when highly reactive volatiles elicit foraging responses at low scent
concentrations.

Increased discovery time of flower patches may have pernicious effects on insects. Many 470 insect species rely on flowers for food, mating and oviposition sites, and follow odor trails 471 to locate them (Byers et al., 2014; Chen et al., 2009; Cunningham et al., 2004; Maia et al., 472 2014: Urru et al., 2011). The longer insects must forage for resources, the less efficient they 473 become and the greater risk they face from predators and parasites (e.g., Goodell (2003)). 474 Thus, pollution may reduce insect fitness through changes in food detectability. Such effects 475 however, are likely to be species specific and context specific in severity. Insects following 476 relatively rare scent plumes in their foraging environment may be more detrimentally affected 477 than insects relying on locally common odor plumes. Similarly, insects needing to locate a 478 scent plume only once to complete a life stage, such as ovipositing in a flower just prior to 479 death (Dunn et al., 2014), might be less affected than those that must repeatedly locate 480 flowers, given that the inherent advantage of insects finding a bigger plume target would 481 be compounded by repetition. Additionally, insects that wander throughout the landscape 482 searching for odor plumes each time a resource is needed may be more affected than central-483 place foragers that locate resource patches and return to them repeatedly using spatial 484 memory (Menzel et al., 2005). The severity of the effects may also be mitigated by whether 485 an insect navigates using single compounds or complex plumes as stimuli. Insects tracking 486 single scents will be able to follow a scent trail as long as it occurs above the detection 487 threshold. For insects following a floral plume, however, detectability depends on both the 488 reactivity of the individual components of that plume and the ability of insects to recognize 489 and track the plume despite variation in plume composition. 490

⁴⁹¹ Modifications of floral scent quality and quantity can have broad impacts on plant-⁴⁹² insect interactions, not just insect fitness. The most reactive compounds in our study, ⁴⁹³ β -caryophyllene and β -myrcene, are common components of floral volatiles and play diverse ⁴⁹⁴ ecological roles, such as attracting pollinators, herbivores, and parasitoids of herbivores. If

⁴⁹⁵ mutualist insects such as pollinators are less efficient at finding hosts then visitation to host ⁴⁹⁶ plants may be less frequent and pollination services may be reduced. Similarly, if predators ⁴⁹⁷ and parasites are less able to follow the scent trail released by plants that are attacked by ⁴⁹⁸ herbivores then the ecological services they provide to plants will also be diminished.

Overall, we predict that existing air pollution levels in urban and industrial corridors can have deleterious impacts on insect foraging efficiency and on plant-insect interactions. Levels of ambient pollutants, types of volatiles comprising the scent plume, sensitivity of insect antennae to plume constituents, the abundance of host plants in the environment and the air conditions (windy versus calm) of the location may all differentially impact the extent of the negative ecological impacts.

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⁶⁸⁰ Figure captions

Figure 1. (Top) Configuration of the Large Eddy Simulation domain and distribution of
 flowers releasing scents to attract insects, and (bottom) spatial distribution of scent plumes
 and location foraging insects.

Figure 2. Vertical variations of wind speed (top) and standard deviation of the vertical wind speed (bottom) under the influences of friction velocity (u_*) of 0.1 and 0.2 m per s.

Figure 3. Lifetimes of floral volatiles as a function of reactivity with respect to (top) ozone, (middle) hydroxyl radical, and (bottom) nitrate radical under the the influences of the oxidant levels shown on the insets (right hand side). Labels placed on top of symbols are 1: β -Caryophyllene, 2: β -Ocimene, 3: β -Myrcene, 4: Linalool, and 5: α -Pinene.

Figure 4. Three-dimensional scent plume characteristics for (top) β -myrcene and (bottom) β -caryophyllene under the influences of pollution scenarios (i) Sc0, (ii) Sc3, and (iii) Sc5 and friction velocity values of 0.1 m s⁻¹ and 0.2 m s⁻¹. Iso-surfaces corresponding to gas mixing ratios of 0.6 pptv (blue) and 4.8 pptv (purple) are illustrated.

Figure 5. Horizontal cross sections of (left) β -myrcene and (right) β -caryophyllene timeaveraged plumes for pollution scenarios (i) Sc0, (ii) Sc3, and (iii) Sc5. The friction velocity of 0.1 m per s was used in the LES. The cross section is at 1.0 m above the surface.

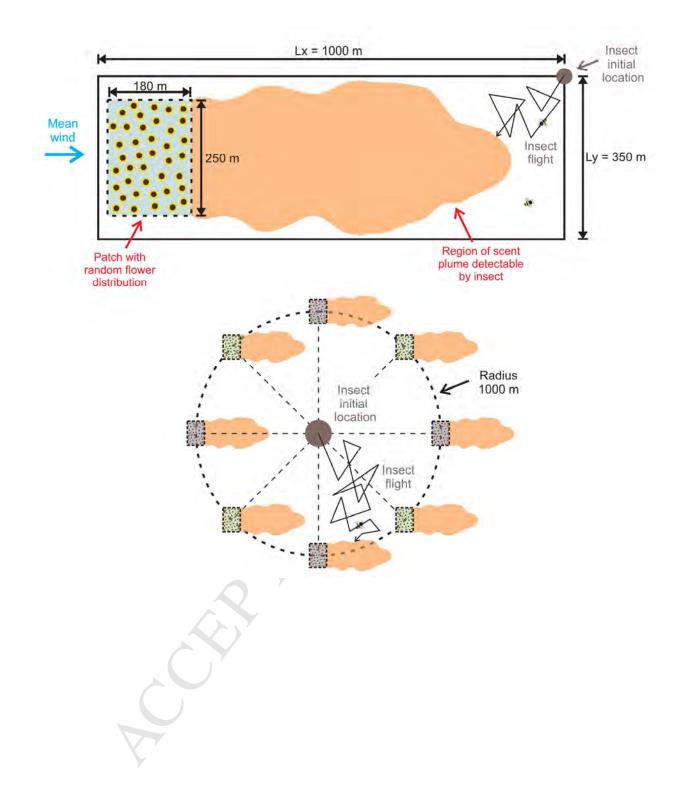
Figure 6. Horizontal cross sections of (left) β -myrcene and (right) β -caryophyllene 697 instantaneous plumes for pollution scenarios (i) Sc0, (ii) Sc3, and (iii) Sc5. The friction 698 velocity of 0.1 m per s was used in the LES. The cross section is at 1.0 m above the surface. 699 **Figure 7.** Fraction of (a) β -myrcene and (b) β -carvophyllene remaining as a function 700 of downwind distance traveled from the source after reacting with hydroxyl radicals, ozone, 701 and nitrate radicals for the four scenarios considered in the modeling study using friction 702 velocity values of 0.1 m per s and 0.2 m per s. The values are averaged at 1.0 m above the 703 surface. 704

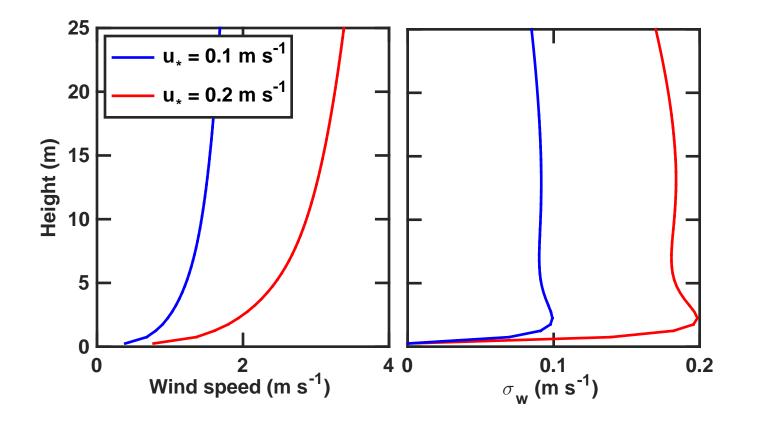
Figure 8. Modification of plume composition as compounds travel and react from sources to downwind areas for (i) no pollution, (ii) 60 ppb of ozone, and (iii) 120 ppb of ozone using u_* of 0.1 m per s. (B) Modification of scents as compounds travel and react from sources to

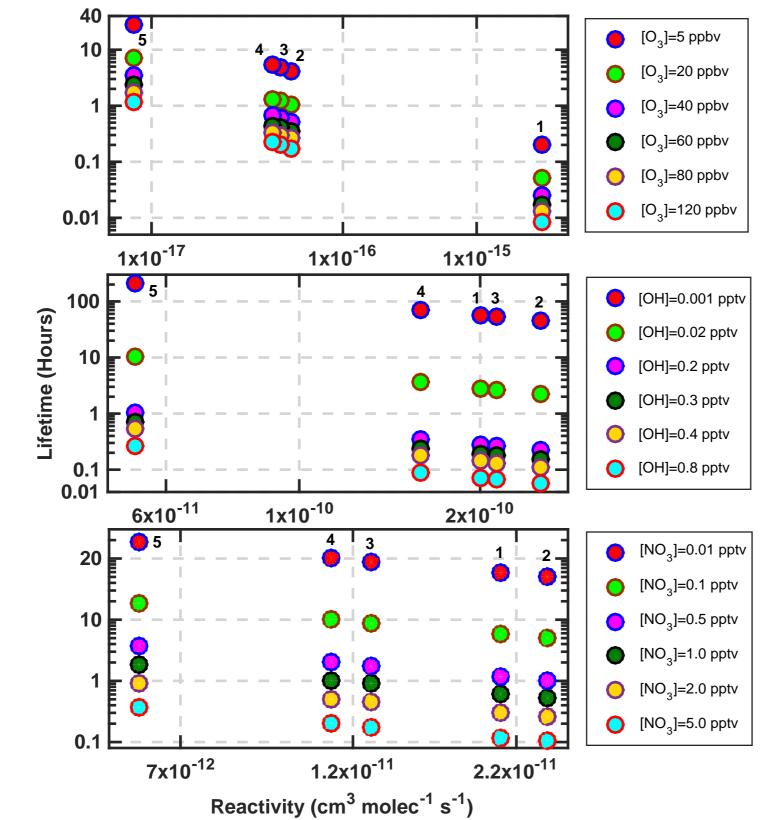
downwind areas for (i) no pollution, (ii) 60 ppb of ozone, and (iii) 120 ppb of ozone using u_* of 0.2 m per s. The values are averaged at 1.0 m above the surface.

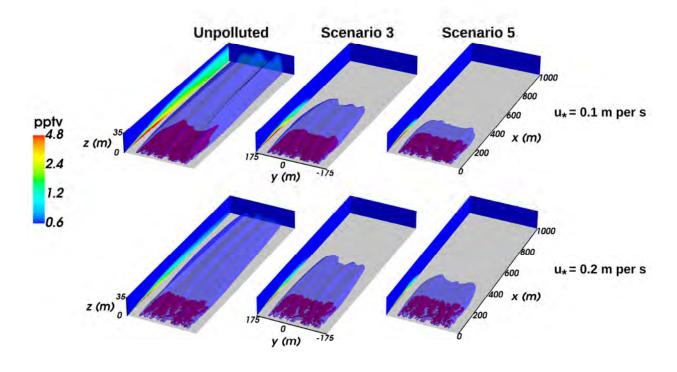
Figure 9. Cumulative distribution function of the foraging times required for insects to locate scent plumes in a landscape with scarce resources (solid lines) and rich in resources (dashed lines). Pollution scenarios are represented by different colors: Sc0 (black), Sc1 (red), Sc2 (green), Sc3 (blue), Sc4 (gray), and Sc5 (orange). Each figure (e.g., Figure A) reports the cumulative distribution function of foraging times for insects with a given threshold detection of α -pinene, β -myrcene, and β -caryophyllene.

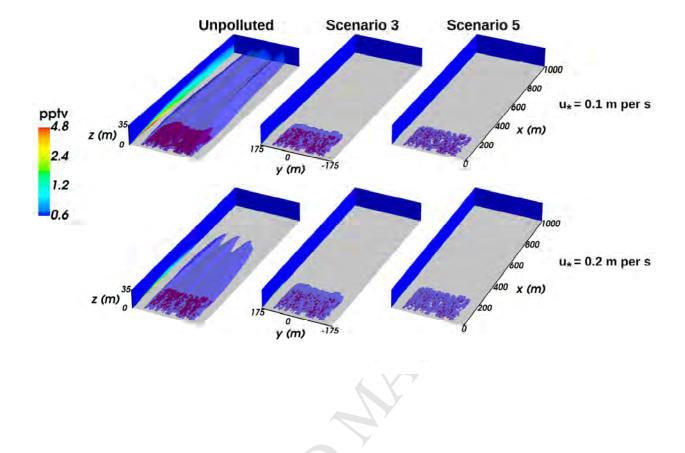
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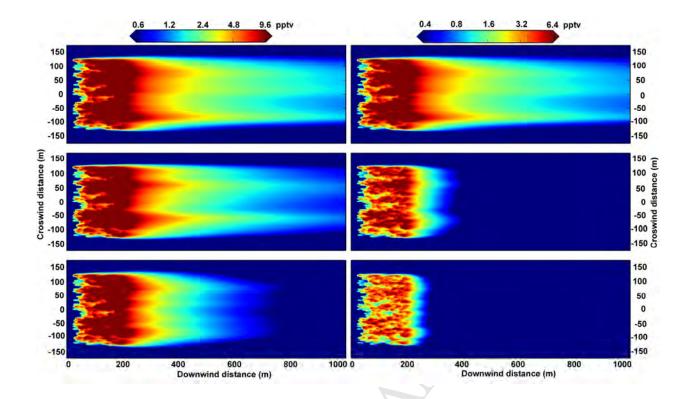


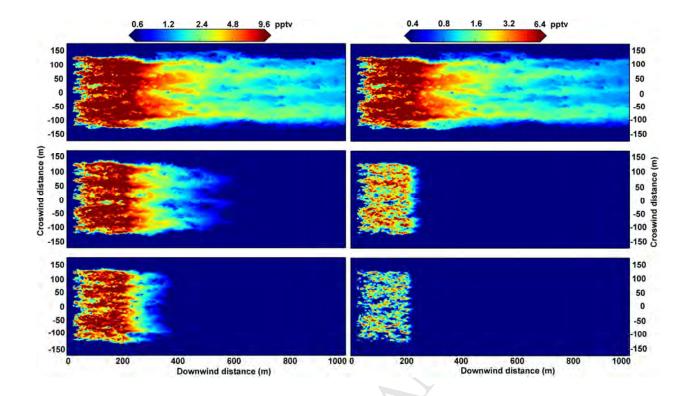


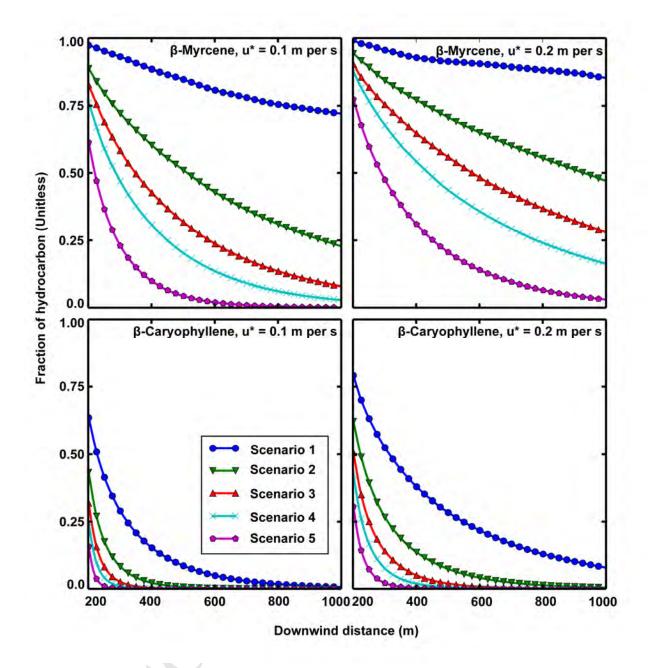


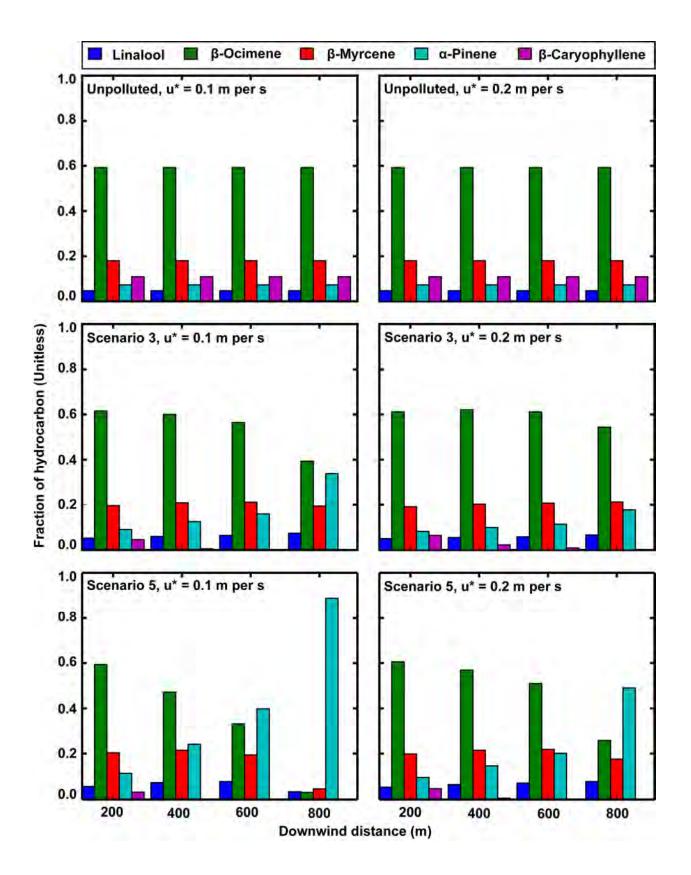


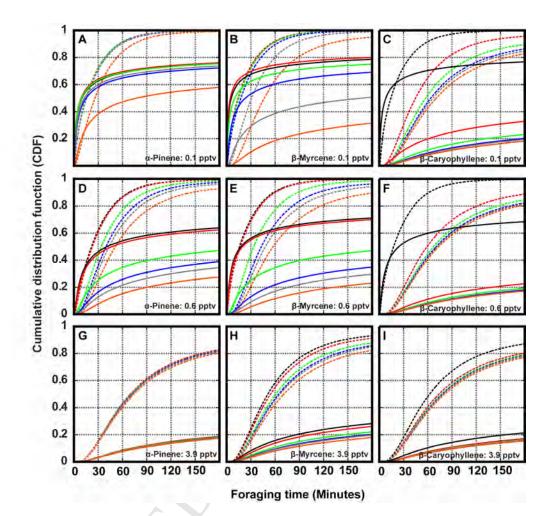












Highlights

- Pollutants such as ozone, nitrate radical, and hydroxyl radical react with floral scents, modifying their chemical composition downwind of sources.
- Ozone mixing ratios greater than 60 parts per billion on a per volume basis can substantially reduce the downwind footprint of floral scents, depending on the reactivity of the chemical constituents of the floral plume.
- Insects increase the time to encounter target floral plumes in polluted air masses due to the modified chemical composition of floral scents.
- Plant-pollinator interactions could be sensitive to changes in floral scent composition, especially if insects are unable to adapt to the modified scentscape.